

## Certificate of Analysis

**Sample Name:** Puariki Instant  
**Description:** Dehydrated aqueous kava root extract

**Batch Number & Sample ID:** B1SL  
**Analysis Identification Code:** 240930U26

**Process/Mix Date:** 26/09/2024  
**Micro Lab Sampling Date:** 27/09/2024  
**ASE Processing Date:** 30/09/2024  
**UHPLC Injection Date:** 30/09/2024  
**Date of Report:** 25/11/2024

**Plant Morphologist:** Joses Laau  
**Chromatographer:** John McGowan  
**Microbiologist:** Ariane Urriza  
**Sample Preparation Technician:** Eva David Livo  
**Quality Control Officer:** Anabel Belen  
**Quality Assurance Manager:** Dianne Manley

### Executive Summary

Total extracted major kavalactones = **7.95%** of sample mass (w/w)  
 Chemotype = **423165**                      Moisture Content: **2.36%** (w/w)  
 Contamination = **Pass**                      Categorisation = **Noble kava**

### K and DHK Ratios

K to DHK: 1.60	DHK to K: 0.63
K to Y: 2.33	DHK to Y: 1.46
K to DMY: 3.60	DHK to DMY: 2.26
K to M: 4.65	DHK to M: 2.92
K to DHM: 5.10	DHK to DHM: 3.20

### Chemical Analysis:

**Sample Preparation:** 1.000 g processed kava powder dispersed with silica sand to fill 10 mL Dionex™ ASE™ cell

**Extraction method:** Accelerated Solvent Extraction (ASE)

**Extraction Process Automation:** Thermo Scientific Dionex™ ASE™ 350 Accelerated Solvent Extractor

**ASE Conditions – Solvent:** HPLC grade Acetonitrile (ACN), **Temperature:** 60 °C, **Pressure:** 105 Bar, **Pre-incubation:** 5 min,

**Static Hold:** 20 min, **Rinse Volume:** 150%, **Dilution to Working Range:** 1 part ASE™ filtrate + 9 parts solvent (ACN) to give 1/10

**Pre-UHPLC Particulate Exclusion:** ASE™ filtrate passed through Dionex™ D28 cellulose filter prior to dilution to working concentration, then passed through 0.22 µm hydrophilic PTFE filter prior to injection

### Chromatographic Conditions:

**System:** Thermo Scientific Vanquish Horizon Ultra-High-Performance Liquid-Chromatography

**Instrument Components:** VF-A10-A Split Sampler, VF-P10-A binary pump, VH-C10-A Column Compartment, and VF-D11-A Diode Array Detector

**Column:** 200 x 2.1 mm Hypersil GOLD, 1.9 µm particle size

**Mobile Phase:** 5% isopropanol to 97% isopropanol in water (nonlinear). Total runtime 15.9 minutes

**Column Temperature:** 60 °C, with active pre-heating      **Injection Volume:** 2.00 µL      **Organic Modifier:** None

**UV Detection:** 362 nm (aflatoxin B<sub>1</sub> and B<sub>2</sub> identification), 341 nm (flavokavain and aflatoxin G<sub>1</sub> and G<sub>2</sub> identification), 246 nm (kavalactone identification), and 218 nm (kavalactone and aflatoxin secondary peaks); Peak identification assisted by elution time and spectrum matching. Relative quantification calculations based on channel 3 (246 nm)

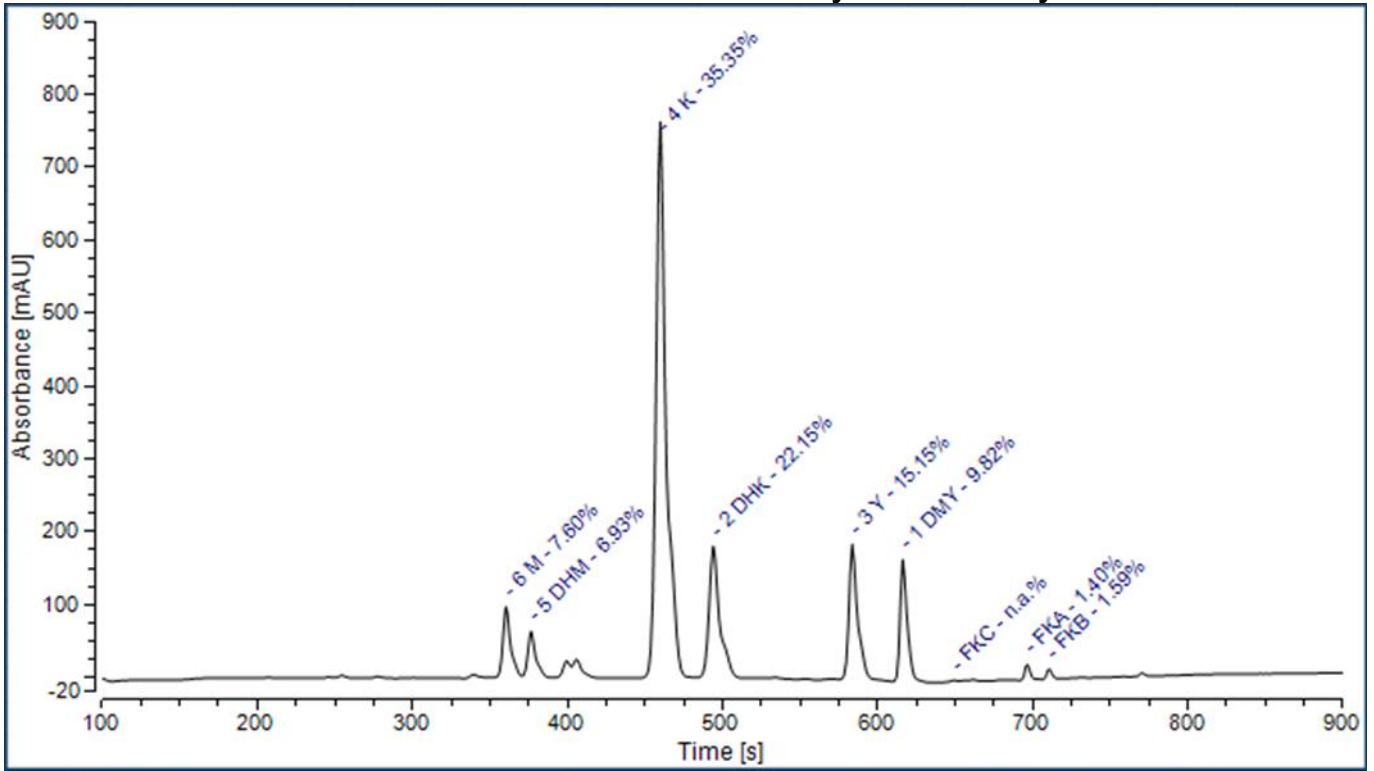
**Calibration Standards:** Correlation coefficient for all identified compounds is greater than 99.995% on a 20-point calibration curve derived by serial dilution of 1 ampoule of Cerilliant (kavalactones and flavokavains) and 1 ampoule of Ehrenstorfer (aflatoxins) certified analytical reference standards. Analytical balances calibrated with certified class OIML E2 weights with uncertainty +/- 0.000016 g (NATA accredited for compliance with ISO/IEC 17025, by laboratory No.3279)

**Instrument Method:** **Software:** Chromeleon 7.2.10 **Program:** 240604 Hypersil iPrOH

**Processing Method:** **Software:** Chromeleon 7.3.2 **Program:** 240606 Hypersil iPrOH Pro

**Confidence Probability:** Lower = 99.5%      Upper = 99.5%

## Determination of Relative Kavalactone Content by UHPLC Analysis at 246 nm



Peak labelled percentages represent that compound's abundance relative to the total amount of quantified compounds in the sample

Integration Results													
Chem #	Rel.Amt	Spectrum	Rtn T.	Rel.Ar	Rel.Ht	R <sup>2</sup>	HV LoD	Cor.Coeff	Cal.	Lower	Amount	Upper	Extracted
Name abv	%	Match	min	%	%	%	(mg/kg)	%	Pts.	Limit	(mg/kg)	Limit	% of mass
4 K	35.355	999.987	7.664	55.45	50.96	99.997	2.9065	99.998	20	28884	28978	29073	2.898
2 DHK	22.154	999.560	8.235	13.72	12.08	99.998	2.2489	99.999	20	18114	18158	18203	1.816
3 Y	15.151	999.689	9.728	11.75	12.46	99.999	2.0071	99.999	20	12390	12418	12446	1.242
1 DMY	9.820	999.151	10.271	8.62	11.19	99.998	2.2349	99.999	20	8024	8049	8074	0.805
6 M	7.599	998.980	6.007	5.40	6.50	99.998	2.3968	99.999	20	6203	6228	6254	0.623
5 DHM	6.934	995.831	6.275	3.63	4.25	99.998	2.0995	99.999	20	5661	5683	5706	0.568
FKCB	1.586	999.292	11.841	0.55	0.98	99.991	0.4956	99.996	20	1293	1300	1307	0.130
FKA	1.402	992.655	11.605	0.83	1.48	99.994	0.4032	99.997	20	1144	1149	1155	0.115
FKC	n.a.	957.267	10.820	0.05	0.09	99.999	0.1742	99.999	20	n.a.	n.a.	n.a.	n.a.
<b>Totals:</b>	<b>100%</b>			<b>100%</b>	<b>100%</b>	<b>Major Kavalactones Extracted (% of total mass):</b>							<b>7.952</b>
<b>Flavokavains Extracted (relative %):</b>				<b>2.988</b>				<b>Flavokavains Extracted (% of total mass):</b>				<b>0.24</b>	
Peak Results													
Peak Name	Peak No.	Purity Match	Ret. T. (S)	Signal to Noise	Peak to Valley	Area mAU*min	Height mAU	Width 50% min	Type	Res (EP)	Asym (EP)	Plates (EP)	
4 K	3	999	459.842	51.4	1373.27	95.917	762.714	0.17	BM *	3.31	1.42	32598	
2 DHK	4	997	494.082	12.2	325.51	23.728	180.788	0.18	MB*	9.20	1.46	35059	
3 Y	5	998	583.662	174.5	n.a.	20.319	186.453	0.15	BMB	3.85	1.50	67684	
1 DMY	6	1000	616.262	156.8	n.a.	14.911	167.537	0.13	BMB	4.63	1.43	94644	
6 M	1	999	360.402	6.6	47.97	9.347	97.225	0.13	BM *	2.00	1.41	32328	
5 DHM	2	999	376.522	4.3	31.38	6.282	63.594	0.14	MB*	9.10	1.38	33968	
FKCB	9	996	710.482	11.4	58.29	0.946	14.741	0.10	MB	n.a.	1.34	231557	
FKA	8	990	696.302	17.1	87.65	1.439	22.168	0.10	BM	2.39	1.32	214850	
FKC	7	901	649.222	1.1	n.a.	0.086	1.409	0.13	BMB*	7.69	1.05	171987	

Disclaimer: The testing protocols employed utilise samples and are representative only of the respective batch, not necessarily other batches or products, even if apparently identical.

These analytical tests have been conducted by suitably qualified personnel on reputable equipment, using high-quality reagents and robust protocols, based upon industry standards. The results are generated in-house, and we believe them to be accurate and precise, however, despite our best efforts, errors may exist; No guarantee is expressed or implied.

These results should not be used as a final determination for use in a finished product; It is recommended that they be verified by the purchaser's quality control department and through the third-party services of an additional certified testing laboratory to ensure the purchased material meets specifications.

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## Summary of Microbial Analysis:

Microbial analyses are carried out in accordance with Forney Enterprises' Quality Assurance Programme. Analyses are performed in our modern, well-equipped, built-for-purpose microbiology laboratory, by experienced staff who are skilled in the art, using aseptic technique, calibrated equipment, and high-quality reagents, and incorporate the use of controls. Any given test may be performed using more than one method, substrate, or growth medium, including (but not limited to) 3M Petrifilm, HyServe Compact Dry plates, traditional and chromogenic agars, and culture-specific broths to validate results. We combine the use of industry standard protocols (such as FDA BAM and AOAC OMA) and proprietary methods developed in-house, however, the results reported are from the most sensitive method used (highest test counts). Forney kava is produced in a closely regulated HACCP certified facility, with continuous environmental monitoring and comprehensive testing throughout the production process. The figures below result from testing the finished product as packaged. Kava powder which does not meet the strictest criteria cannot bear the Forney logo.

Indicator Organism	Test Results (cfu/g)
Aerobic Plate Count (TPC)	550
Coliform	None detected
<i>Escherichia coli</i>	None detected
Yeast	None detected
Mould	None detected
<i>Staphylococcus spp.</i>	None detected
<i>Salmonella spp.</i>	None detected
<i>Listeria spp.</i>	None detected